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U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY'S DOCKET NUMBER

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U.S. APPLICATION NO. (If known, see 37 CFR 1.5)

09/19/396

TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371INTERNATIONAL APPLICATION NO.
PCT/SE98/00555INTERNATIONAL FILING DATE
26 MARCH 1998PRIORITY DATE CLAIMED
26 MARCH 1997

TITLE OF INVENTION

ANTIGENIC FUSIONPROTEIN CARRYING GAL α 1,3GAL EPITOPES

APPLICANT(S) FOR DO/EO/US

Jan HOLGERSSON and Jining LIU

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
4. ☐ A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
5. ☒ A copy of the International Application as filed (35 U.S.C. 371(c)(2))
 - a. ☒ is transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☐ has been transmitted by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☒ A translation of the International Application into English (35 U.S.C. 371(c)(2)).
7. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))
 - a. ☐ are transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☐ have been transmitted by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☐ have not been made and will not be made.
8. ☒ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☒ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).
10. ☒ A translation of the annexes of the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

Items 11. to 16. below concern document(s) or information included:

11. ☒ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
12. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. ☒ A **FIRST** preliminary amendment.
☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
14. ☐ A substitute specification.
15. ☐ A change of power of attorney and/or address letter.
16. ☒ Other items or information: **International Search Report (PCT/ISA/210)**
Patent Application Data Entry Format Sheets

17. ☒ The following fees are submitted:

BASIC NATIONAL FEE (37 CFR 1.492 (a) (1) - (5)):

Neither international preliminary examination fee (37 CFR 1.482)
nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO
and International Search Report not prepared by the EPO or JPO \$ 970.00

International preliminary examination fee (37 CFR 1.482) not paid to
USPTO but International Search Report prepared by the EPO or JPO \$840.00

International preliminary examination fee (37 CFR 1.482) not paid to USPTO
but international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$760.00

International preliminary examination fee (37 CFR 1.482) paid to USPTO
but all claims did not satisfy provisions of PCT Article 33(1)-(4) \$670.00

International preliminary examination fee (37 CFR 1.482) paid to USPTO
and all claims satisfied provisions of PCT Article 33(1)-(4) \$96.00

ENTER APPROPRIATE BASIC FEE AMOUNT =

\$ 970

Surcharge of \$130.00 for furnishing the oath or declaration later than ☒ 20 ☐ 30
months from the earliest claimed priority date (37 CFR 1.492(e)).

\$ 130

CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE	\$
Total claims	20 - 20 =	0	x \$18.00	\$
Independent claims	1 - 3 =	0	x \$78.00	\$
MULTIPLE DEPENDENT CLAIM(S) (if applicable)			+ \$260.00	\$

TOTAL OF ABOVE CALCULATIONS =

\$1,100

Reduction of 1/2 for filing by small entity, if applicable. A Small Entity Statement
must also be filed (Note 37 CFR 1.9, 1.27, 1.28). + \$

SUBTOTAL =

\$1,100

Processing fee of \$130.00 for furnishing the English translation later than ☐ 20 ☐ 30
months from the earliest claimed priority date (37 CFR 1.492(f)). \$

TOTAL NATIONAL FEE =

\$1,100

Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be
accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property + \$

TOTAL FEES ENCLOSED =

\$1,100

Amount to be

refunded:

\$

charged:

\$

- a. ☒ A check in the amount of \$ 1,100 to cover the above fees is enclosed.
- b. ☐ Please charge my Deposit Account No. _____ in the amount of \$ _____ to cover the above fees.
A duplicate copy of this sheet is enclosed.
- c. ☒ The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any
overpayment to Deposit Account No. 25-0120. A duplicate copy of this sheet is enclosed.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR
1.137 (a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO:

November 25, 1998

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SIGNATURE

Benoit Castel

NAME

35, 041

REGISTRATION NUMBER

PATENTS

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of

Jan HOLGERSSON et al.

Serial No. (unknown)

Filed herewith

ANTIGENIC FUSIONPROTEIN
CARRYING GAL α 1,3GAL
EPITOPES

PRELIMINARY AMENDMENT

Assistant Commissioner for Patents

Washington, D.C. 20231

Sir:

Prior to calculation of the filing fee, please
amend the above-identified application as follows:

IN THE CLAIMS:

Claim 3, line 1, change "any one of claims 1 and
2," to --claim 1,--.

Claim 4, line 1, change "any one of the preceding
claims," to --claim 1,--.

Claim 5, line 1, change "any one of the preceding
claims," to --claim 1,--.

Claim 9, line 1, change "any one of the preceding
claims," to --claim 1,--.

Claim 13, line 1, change "any one of claims 9-12,"
to --claim 9,--.

Claim 14, line 1, change "any one of the claims
9-12," to --claim 9,--.

COMMUNICATIONS SECTION

Jan HOLGERSSON et al.

Claim 15, line 2, change "any one of claims 1-14"
to --claim 1,--.

Claim 19, line 3, change "any one of claims 1-14"
to --claim 1,--.

Claim 20, lines 3 and 4, change "any one of claims
1-14" to --claim 1--.

Respectfully submitted,

YOUNG & THOMPSON

By

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November 25, 1998

305 Rec'd PCT/PTO 25 NOV 1998

ANTIGENIC FUSIONPROTEIN CARRYING GAL α 1,3GAL EPITOPES

Technical field

The present invention relates to antigenic fusionproteins carrying multiple Gal α 1,3Gal-epitopes for the removal of foreign antibodies, such as xenoreactive human anti-pig antibodies, by absorption.

Background

Many diseases are today curable only by a transplantation of tissue or an organ, such as a kidney or heart. It is sometimes possible to locate a living donor with immunological markers compatible with the transplant recipient, although organ donation by a living donor involves great risks and possible deleterious health effects for the donor. Without any available living donor, the organ must be obtained from a heart-beating human cadaver of high quality and, again, there must be a good immunological match between the donor and the recipient. The situation today is a steadily increasing demand for human organs suitable for transplantation and the gap between said demand and the availability of organs is likely to grow even wider in view of the continuing improvements made in transplantation procedures and outcome. The most promising possible answer to this problem is xenotransplantation, i.e. transplantation of tissue or organs between different species. For human patients, the pig is considered the most suitable donor species for medical, practical, ethical and economical reasons.

The main problem in xenografting between discordant species, such as pig to human, is the hyperacute rejection (HAR), which leads to a cessation of the blood flow within minutes following a transplantation. Even though other mechanisms of rejection will ensue after HAR, the general belief is that if HAR could be prevented, the patient's immune system may undergo a process of accommodation, whereafter a conventional immunosuppressive regimen could maintain the compatibility of the patient and the xenograft.

The HAR is caused by preformed, natural antibodies in the receiving species reacting with antigens on the endothelium in donor organs, an interaction which leads to complement and endothelial cell activation, thrombosis, extravasation of white blood cells and, eventually, rejection. Pig antigens reacting with human, natural antibodies have turned out to be carbohydrates (7-10); the major one being the Gal α 1,3Gal epitope which is not expressed in old world monkeys, apes and humans due to an inactivation of the α 1,3 galactosyltransferase (GT) (10-12).

Several methods have been proposed for the removal or elimination of xenoreactive antibodies from the blood of a recipient. Bach et al (Xenotransplantation, Eds: Cooper, D.K.C., et al, Springer Verlag, 1991, Chapter 6) proposed the perfusion of the recipients blood through an organ of the proposed donor species prior to transplantation of another, fresh organ, whereby anti-pig antibodies were removed.

Plasmapheresis has also been proposed for a non-specific removal of naturally occurring antibodies, whereby the graft survival is prolonged (e.g. Cairns et al, Rydberg et al). However, conventional plasmapheresis, or plasma exchange, results in loss of blood volume, which in turn may require a volume replacement with pooled preparations of fresh frozen plasma, human albumin, immunoglobulin etc. In addition, coagulation factors, platelets and antithrombotic factors must also be replaced. Such a treatment carries not only the risk of virus transfer, such as HIV, but also the risk of an anaphylactic reaction to foreign substances. Other negative side effects of plasmapheresis are recipient sensitization and activation of the complement and clotting system. Accordingly, plasmapheresis does not appear to be either practical or safe.

Other methods for the removal of xenoreactive antibodies involve non-specific antibody removal. Protein A, a major component of the cell wall of *S. aureus*, has a high affinity for a portion of the Fc-region of sub-classes 1, 2 and 4 of immunoglobulin G (IgG1, IgG2, IgG4) and has been used for the non-specific removal of anti-HLA antibodies from hypersensitized patients in need of kidney transplants. The efficacy of the Protein A column treatment after kidney transplantation have been reported (Dantal J., et al, New England J. Med. 550: 7-14, 1994; Nilsson, I.M. et al, Blood 58: 38-44, 1981; Palmer, A., et al., The Lancet January 7, 1989, pp. 10-12). One essential drawback with the use of a Protein A column technique in the context of xenotransplantation is, however, the fact that only IgG will be removed. Lately, it has been shown that the antibodies involved in HAR during a transplantation from pig to human may involve several other immunoglobulin classes. In addition, the non-specific antibody removal will cause a general deterioration of the patients immune defense, which quite naturally is not desirable during such a process as a transplantation procedure, where the patient is immunosuppressed.

Leventhal et al (WO 95/31209) propose a method of preventing or ameliorating a hyperacute reaction occurring after transplantation of a pig organ to a primate recipient, including a human. The method involves passing the recipients plasma over a

column with a coupled protein, which binds to and thereby removes immunoglobulin therefrom. The protein is selected from a group consisting of Staphylococcus aureus protein A, Streptococcus protein G and anti-human immunoglobulin antibodies. This method suffers from the same drawbacks as depicted above for the protein A column.

It has been shown (Platt et al, Good et al, Holgersson et al) that pig antigens reacting with human, natural antibodies are carbohydrates, the major one being the Gal α 1,3Gal epitope, which is not expressed in old world monkeys, apes and humans due to an inactivation of the α 1,3 galactosyltransferase.

Recently, McKenzie et al showed that COS cells transfected with the α 1,3-galactosyltransferase cDNA expressed the Gal α 1,3Gal-epitope on their surfaces and could absorb most of the human anti-pig activity from human serum.

Further, Gal α 1,3Gal-derivatized columns have been used to specifically remove anti-pig activity from human serum (22), free Gal α 1,3Gal disaccharides have been shown to prevent binding of anti-pig antibodies to porcine cells, including endothelium (23), and so has porcine stomach mucin (24). However, organic synthesis of saccharides is a very laborious and expensive method, which in addition is rather slow, and accordingly, has not found any wide spread applicability.

Apart from the HAR, xenografts are still typically rejected within days in a process that has been termed delayed xenograft rejection (DXR) (29). DXR is characterized by mononuclear cell activation and graft infiltration, as well as cytokine production (29). The importance of the Gal α 1,3Gal epitope for these cellular events is not known, although human anti-Gal α 1,3Gal antibodies were recently shown to be involved in antibody-dependent cellular cytotoxicity (ADCC) of porcine cells (30).

Thus, there is still a need of cheaper and more efficient methods for the elimination of foreign antibodies, such as pig-antibodies, from the blood from a recipient who is to obtain a xenotransplant. In addition, within the field of xenotransplantation, of methods for the prevention of DXR.

Summary of the invention

The objective of the present invention is to fulfill the above defined need. Accordingly, the present invention provides an antigenic fusionprotein, which carries multiple Gal α 1,3Gal epitopes. In a preferred embodiment, the fusionprotein accor-

ding to the present invention is comprised of a heavily glycosylated mucin part, which may mediate binding to the selectins, and a part conferring immunoglobulin properties. The fusionprotein according to the invention carries a multitude of Gal α 1,3Gal epitopes, that effectively absorbs foreign antibodies, such as anti-pig antibodies, involved in antibody-dependent, complement-mediated killing and ADCC of endothelial cells, such as porcine cells.

Detailed description of the invention

The present invention relates to an antigenic fusionprotein, which carries multiple Gal α 1,3Gal epitopes.

Thus, the antigenic fusionprotein according to the present invention is capable of binding preformed antibodies as well as antibodies produced as a response to a transplanted tissue or organ originating from the species in which the Gal α 1,3 epitopes are expressed, said species preferably being a species different from the antibody producing species. In a preferred embodiment, the antibody producing species is a human being producing antibodies against a foreign transplant, e.g. a pig organ, in which case the Gal α 1,3 epitopes are synthesized by a α 1,3 galactosyltransferase of pig origin. The transplant may be an organ such as a liver, a kidney, a heart etc, or tissue thereof. Accordingly, in the most preferred embodiment of the fusionprotein according to the invention, the antigenic fusionprotein carries multiple Gal α 1,3Gal epitopes synthesized by a α 1,3 galactosyltransferase derived from a porcine species.

Thus, as the fusionprotein according to the present invention may be prepared by the culture of genetically manipulated cells, such as COS cells, it is both cheaper and easier to produce than the previously saccharides produced by organic synthesis. Even though a COS cell expressing the Gal α 1,3Gal epitope on its surface has been described (12), the present invention is the first proposal of a recombinant fusionprotein, which carries multiple such epitopes. The fusionprotein according to the invention can easily be designed to include other peptides and parts, which may be advantageous for a particular application. Examples of other components of the fusionprotein according to the invention will be described more detailed below.

Thus, in a preferred embodiment, the antigenic fusionprotein according to the invention further comprises a part, which mediates binding to selectin, such as P-selectin. Said part is preferably a highly glycosylated protein, such as a protein of mucin type. The mucins are due to their high content of O-linked carbohydrates especially advantageous together with the Gal α 1,3Gal epitope in the fusionprotein

according to the invention, as the antigenic properties in the present context thereby are greatly improved. Thus, it has been shown that the binding of antibodies which are reactive with the Gal α 1,3Gal epitope is even more efficient if said epitope is presented by a protein of mucin type, which indicates that the binding in fact may involve more than said epitope alone.

In the hitherto most preferred embodiment of the fusionprotein according to the invention, the part that mediates binding to selectin is the P-selectin glycoprotein ligand-1 (PSGL-1) or an essential part thereof. However, other cell membrane-anchored proteins containing mucin-type domains have been characterized and may be used in the fusionprotein according to the invention as appropriate, such as CD34, CD43, GlyCAM-1, PSGL-1, MAdCAM, CD96, CD45 and RBC glycophorins. In the experimental part of the present application, an example wherein said P-selectin glycoprotein ligand-1 (PSGL-1) is derived from HL-60 cells, is shown. Theoretically, the anti-pig antibody repertoire may recognize the Gal α 1,3Gal epitope in various structural contexts determined by the core saccharide presenting the epitope, neighbouring branching points, and the proximity to other sugar residues such as fucose and sialic acid (31-33). If Gal α 1,3Gal disaccharides or Gal α 1,3Gal β 1,4GlcNAc trisaccharides are used as absorbers some specificities of the repertoire might not be efficiently absorbed.

The properties of the part of the fusionprotein mediating selectin binding according to the invention as well as further reasoning concerning the choice thereof is further discussed below, see the section "Discussion".

In an especially advantageous embodiment, the antigenic fusionprotein according to the present invention further comprises a part which confers immunoglobulin properties. The immunoglobulin parts are advantageous for the design of an efficient and simple method of coupling the fusionprotein according to the invention to a solid support to be used for purifying plasma from a recipient of a xenotransplant from antibodies against said transplant. An immunoglobulin part can also be included in the fusion protein according to the invention in the preferred case, where the fusionprotein is produced in a cell which secretes it, whereafter the immunoglobulin part is used for the purification of said secreted fusionprotein from the culture.

Thus, according to an advantageous embodiment of the antigenic fusionprotein according to the present invention, the part that confers immunoglobulin properties is an immunoglobulin or a part thereof, such as IgG or a part thereof. Preferably, said

part that confers immunoglobulin properties is the Fc part of an immunoglobulin, preferably of IgG, or an essential part thereof. In a particular embodiment of the present invention, said immunoglobulin property conferring part is IgG_{2b}, preferably the Fc part thereof. In an example of a fusionprotein according to the present invention, said part that confers immunoglobulin properties is of non-human origin, and is preferably derived from mouse. However, for certain applications said part might more preferably be of human origin.

The fusionprotein according to the present invention is preferably a recombinant fusionprotein. It may have been produced in a recombinant cell line, preferably an eukaryotic cell line, *e.g.* a COS cell line, following cotransfection of the cDNA for the mucin/immunoglobulin fusion protein and the cDNA for the porcine α 1,3 galactosyltransferase.

The fusionprotein according to the present invention may for example be used as an absorber for elimination of antibodies from blood plasma. The use is discussed in more detail below, in the section "Discussion".

Another aspect of the present invention is cDNA molecule, which comprises a cDNA sequence coding for a fusionprotein as defined above or a derivative or variant thereof.

Yet another aspect of the present invention is a vector, which comprises the cDNA molecule as described above together with appropriate control sequences, such as primers etc. The skilled in the art can easily choose appropriate elements for this end.

Another aspect of the present invention is a cell line transfected with the above defined vector. The cell line is preferably eukaryotic, *e.g.* a COS cell line. In the preferred embodiment said cells are secreting the fusionprotein according to the invention into the culture medium, which makes the recovery thereof easier and, accordingly, cheaper than corresponding methods for synthesis thereof would be.

Another aspect of the invention is an absorber comprised of a fusionprotein according to the present invention coupled to a solid support. The absorber according to the invention is used in a pretransplant extracorporeal immunoabsorption set up to remove anti-pig antibodies involved in antibody dependent, complement- as well as cell-mediated cytotoxicity of pig endothelial cells.

Finally, a last aspect of the invention is a method of purifying blood plasma from foreign antibodies, e.g. elimination of anti-pig antibodies from human blood plasma. The method involves withdrawal of plasma from a patient, such as a patient who is to receive a transplant of porcine origin, bringing said plasma in contact with a fusionprotein according to the invention to bind anti-pig antibodies thereto, whereby the anti-pig antibodies are eliminated from the plasma, and thereafter reinfusing the plasma into the patient. The method according to the invention has been shown to be more efficient than the previously known methods in the prevention of HAR, possibly thanks to the large amount of carbohydrate in the fusionprotein. Indeed, it is feasible that the present method also contributes in a prevention of delayed xenograft rejection (DXR), as the spectra of antibodies eliminated by the fusionprotein according to the invention presumably is broader than in the prior methods.

EXPERIMENTAL

In the present application, the following abbreviations are used:

ADCC, antibody-dependent cellular cytotoxicity; BSA, bovine serum albumin; DXR, delayed xenorejection; ELISA, enzyme-linked immunosorbent assay; FT, fucosyltransferase; Gal, D-galactose; GT, galactosyltransferase; Glc, D-glucose; GlcNAc, D-N-acetylglucosamine; GlyCAM-1, glycosylation-dependent cell adhesion molecule-1; HAR, hyperacute rejection; Ig, immunoglobulin; MAdCAM, mucosal addressin cell adhesion molecule; PAEC, porcine aortic endothelial cells; PBMC, peripheral blood mononuclear cells; PSGL-1, P-selectin glycoprotein ligand-1; RBC, red blood cell; SDS-PAGE, sodium dodecyl sulphate - polyacrylamide gel electrophoresis

Materials and Methods

Cell culture. COS-7 m6 cells (35) and the SV40 Large T antigen immortalized porcine aortic endothelial cell line, PEC-A (36), were passaged in Dulbecco's modified Eagle's medium (DMEM), with 10% fetal bovine serum (FBS) and 25 µg/ml gentamicin sulfate. The human erythroleukemic cell line, K562, and the Burkitt's lymphoma cell line, Raji, were obtained from ATCC and cultured in HEPES-buffered RPMI 1640 with 10% FBS, 100 IU/ml penicillin and 100 µg/ml streptomycin.

Construction of expression vectors. The porcine α 1,3 GT (37-39) was PCR amplified off pig spleen cDNA using a forward primer having six codons of complementarity to the 5' end of the coding sequence, a Kozak translational initiation consensus sequence and a Hind3 restriction site, and a reverse primer with six codons of

complementarity to the 3' end of the coding sequence, a translational stop and a NotI restriction site. The amplified α 1,3GT cDNA was cloned into the polylinker of CDM8 using Hind3 and NotI (35). The P-selectin glycoprotein ligand-1 (PSGL-1) - a highly glycosylated mucin-type protein mediating binding to P-selectin (40) - coding sequence was obtained by PCR off an HL-60 cDNA library, cloned into CDM8 with Hind3 and NotI, and confirmed by DNA sequencing. The mucin/immunoglobulin expression plasmid was constructed by fusing the PCR-amplified cDNA of the extracellular part of PSGL-1 in frame via a BamHI site, to the Fc part (hinge, CH2 and CH3) of mouse IgG_{2b} carried as an expression cassette in CDM7 (Seed, B. *et al*, unpublished).

Production and purification of secreted mucin/immunoglobulin chimeras. COS m6 cell were transfected using the DEAE-dextran protocol and 1 μ g of CsCl-gradient purified plasmid DNA per ml transfection cocktail. COS cells were transfected at approximately 70 % confluency with empty vector (CDM8), the PSGL1/mIgG_{2b} plasmid alone or in combination with the α 1,3GT encoding plasmid. Transfected cells were trypsinized and transferred to new flasks the day after transfection. Following adherence for approximately 12 hrs, the medium was discarded, the cells washed with phosphate buffered saline (PBS), and subsequently incubated another 7 days in serum-free, AIM-V medium (cat.nr. 12030, Life technologies Inc.). After incubation, supernatants were collected, debris spun down (1400 x g, 20 minutes), and NaN₃ added to 0.02%. PSGL1/mIgG_{2b} fusion protein was purified on goat anti-mouse IgG agarose beads (A-6531, Sigma) by rolling head over tail, over night at 4 °C. The beads were washed in PBS and subsequently used for SDS-PAGE and Western blot analysis, or for absorption of human AB serum and purified human immunoglobulins.

Purification of human IgG, IgM and IgA. Human IgG, IgM and IgA were purified from human AB serum - pooled from more than 20 healthy blood donors - using goat anti-human IgG (Fc specific; A-3316, Sigma), IgM (μ -chain specific; A-9935, Sigma), and IgA (α -chain specific; A-2691, Sigma) agarose beads. Briefly, 5 ml of slurry (2.5 ml packed beads) were poured into a column of 10 mm diameter and washed with PBS. Ten milliliter of human pooled AB serum was applied at 1 ml/minute using a peristaltic pump, washed with several column volumes of PBS, and eluted with 0.1 M glycine, 0.15 M NaCl, pH 2.4 using a flow rate of 1 ml/minute. One milliliter fractions were collected in tubes containing 0.7 ml of neutralizing buffer (0.2 M Tris/HCl, pH 9). The absorption at 280 nm was read spectrophotometrically and tubes containing protein were pooled. dialyzed against 1%

PBS, and lyophilized. Lyophilized immunoglobulins were resuspended in distilled water and the concentrations adjusted to 16 mg/ml for IgG, 4 mg/ml for IgA and 2 mg/ml for IgM.

SDS-PAGE and Western blotting. SDS-PAGE was run by the method of Leammli with a 5 % stacking gel and a 6 or 10 % resolving gel using a vertical Mini-PROTEAN II electrophoresis system (Bio-Rad, Hercules, CA) (41). Separated proteins were electrophoretically blotted onto Hybond™-C extra membranes (Amersham) using a Mini Trans-Blot electrophoretic transfer cell (Bio-Rad, Hercules, CA) (42). Protein gels were stained using a silver staining kit according to the manufacturer's instructions (Bio-Rad, Hercules, CA). Following blocking for at least 2 hrs in 3 % BSA in PBS, the membranes were probed for 2 hrs in room temperature with peroxidase-conjugated *Bandereia simplicifolia* isolectin B₄ (L-5391, Sigma) diluted to a concentration of 1 µg/ml in PBS, pH 6.8 containing 0.2 mM CaCl₂. The membranes were washed 5 times with PBS, pH 6.8, and bound lectin was visualized by chemiluminescens using the ECL™ kit according to the instructions of the manufacturer (Amersham).

Quantification of PSGL1/mIgG_{2b} by anti-mouse IgG Fc ELISA. The concentration of fusion protein in cell culture supernatants before and after absorption was determined by a 96-well ELISA assay, in which fusion proteins were captured with an affinity purified, polyclonal goat anti-mouse IgG Fc antibody (cat.nr. 55482, Cappel/Organon Teknika, Durham, NC). Following blocking with 3 % BSA in PBS, the fusion proteins were captured and detected with a peroxidase-conjugated, affinity purified, polyclonal anti-mouse IgG Fc antibody (cat.nr. 55566, Organon Teknika, Durham, NC) using O-phenylenediamine dihydrochloride as substrate (Sigma). The plate was read at 492 nm and the ELISA calibrated using a dilution series of purified mouse IgG Fc fragments (cat.nr. 015-000-008, Jackson Immuno-Research Labs., Inc., West Grove, PA) resuspended in AIM V serum-free medium.

Porcine aortic endothelial cell ELISA. PEC-A cells were seeded at a density of 15 000 cells/well in gelatin-coated 96-well plates (Nunc, Denmark) and cultured for 48 hrs in AIM V serum-free medium. The plate was washed 5 times in 0.15 M NaCl containing 0.02 % Tween 20 and incubated for 1 hr in room temperature with 50 µl/well of purified human IgG, IgM, and IgA in PBS, starting at a concentration of 8, 1, and 2 mg/ml, respectively. The plate was washed again as above, and 50 µl of alkaline phosphatase-conjugated goat anti-human IgG (γ-chain specific; A3312, Sigma), IgM (µ-chain specific; A1067, Sigma) and IgA (α-chain specific; A3062,

Sigma) F(ab)₂ fragments diluted 1:200 in PBS were added and incubated for 1 hr at room temperature. The plate was washed as above, incubated with the substrate *p*-nitrophenyl phosphate (Sigma 104-105), and read at 405 nm.

5 **Porcine aortic endothelial cell cytotoxicity assay.** PEC-A cells were seeded and cultured in 96-well plates as described for the PEC-A ELISA. Following 48 hrs of culture, the cells were loaded for 1 hr at 37 °C with Na₂⁵¹CrO₄ (cat.nr. CJS4, Amersham), 1 µCi/well, and washed 3 times with AIM V medium. Fifty microliter of serially diluted, absorbed or non-absorbed human AB serum or purified human
10 IgG, IgA, or IgM antibodies were added together with 50 µl of rabbit serum (Cat. no. 439665, Biotest AG, Dreieich, Germany) as a source of complement. Following 4 hrs incubation at 37 °C in a 5 % CO₂ atmosphere, the supernatants were harvested using a Skatron supernatant collection system (Skatro Instruments, Norway) and analyzed in a γ-counter (1282 Compugamma, LKB Wallac). Each serum and Ig
15 sample were analyzed in triplicate. The percent killing was calculated as the measured minus the spontaneous release divided by the difference between the maximum and the spontaneous release.

20 **Antibody-dependent cellular cytotoxicity (ADCC).** Human PBMC were isolated from fresh buffy coats prepared from healthy donors at the Blood bank of the South hospital, Stockholm. Six milliliter of buffy coat was mixed in a 50 ml polypropylene tube with 15 ml of PBS containing 1 mg/ml BSA and 3.35 mg/ml EDTA. Following centrifugation at 500 x g for 10 minutes, the platelet rich supernatant was discarded, and 6 ml of the lower phase was mixed with 6 ml of Hank's balanced salt solution (HBSS), and underlayered with 6 ml of Lymphoprep™ (Nycomed Pharma AS).
25 Following centrifugation (800 x g, 20 min.), the interface was transferred to a new tube, washed three times in HBSS, and resuspended in serum-free AIM V medium. The final step in the effector cell preparation was to transfer the PBMC to tissue culture flasks that were incubated for 1 hr at 37 °C and 5 % CO₂ to remove plastic-adherent cells. Target cells were K562 and Raji cells kept as described above, or
30 PEC-A cells that had been trypsinized the day before the assay and subsequently cultured in AIM V serum-free medium to prevent readhesion to the plastic surface. At the time for the assay the PEC-A cells were washed off the bottom of the flask. Target cells were loaded with Na₂⁵¹CrO₄, 100 µCi/1 x 10⁶ cells, for 1 hr at 37 °C,
35 washed 3 times in HBSS and resuspended in AIM V to a final concentration of 5 x 10⁴/ml. Five thousand target cells were added to each well with effector cells in 200 µl AIM V medium with and without 10 % heat-inactivated, human AB serum at an effector (E):target (T) ratio ranging from 50:1 in two-fold dilutions to 6.25:1.

Spontaneous release was read in wells with 5 000 target cells incubated in 200 μ l AIM V medium without effector cells and maximum release was read in wells where 5 000 target cells in 100 μ l AIM V were incubated with 100 μ l 5 % Triton X-100. Each E:T ratio was analyzed in triplicate. Following incubation at 37 °C for 4 hrs, the supernatants were harvested using a Skatron supernatant collection system (Skatro Instruments, Norway) and analyzed in a γ -counter (LKB Wallac). The percent killing was calculated as the measured minus the spontaneous release divided by the difference between the maximum and the spontaneous release.

RESULTS

Expression and characterization of the PSGL1/mIgG_{2b} fusion protein. Supernatants from COS-7 m6 cells transfected with the vector plasmid CDM8, the PSGL1/mIgG_{2b} plasmid, or the PSGL1/mIgG_{2b} together with the porcine α 1,3 GT plasmid, were collected approximately 7 days after transfection. Secreted mucin/Ig fusion proteins were purified by absorption on anti-mouse IgG agarose beads and subjected to SDS-PAGE and Western blotting using the *Bandereia simplicifolia* isolectin B₄ (BSA IB₄) for detection. As seen in Fig. 1, the fusion protein migrated under reducing conditions as a broad band with an apparent molecular weight of 145 kDa that stained relatively poorly with silver. The heterogeneity in size, approximately 125 to 165 kDa, and poor stainability is in concordance with previous observations with respect to the behavior of highly glycosylated, mucin-type proteins (43, 44). The fusion protein is most likely produced as a homodimer because SDS-PAGE under non-reducing conditions revealed a double-band of an apparent molecular weight of more than 250 kDa. The amounts of fusion protein affinity-purified from the two supernatants derived from the same number of COS cells transfected with the PSGL1/mIgG_{2b} plasmid alone or together with the α 1,3GT plasmid, respectively, were similar. Probing the electroblotted membranes with BSA IB₄ revealed strong staining of the fusion protein obtained following cotransfection with the porcine α 1,3 GT (Fig. 1). It is clear, though, that the PSGL1/mIgG_{2b} fusion protein produced in COS-7 m6 cells without cotransfection of the α 1,3 GT cDNA also exhibited weak staining with the BSA IB₄ lectin, in spite of the fact that COS cells are derived from the Simian monkey - an old world monkey lacking α 1,3 GT activity. This indicates that the BSA IB₄ lectin has a slightly broader specificity than just Gal α 1,3Gal epitopes (45). Nevertheless, cotransfection of the porcine α 1,3GT cDNA supported the expression of a highly Gal α 1,3Gal-substituted PSGL1/mIgG_{2b} fusion protein.

Quantification of PSGL1/mIgG_{2b} chimeras in supernatants of transfected COS cells, and on goat anti-mouse IgG agarose beads following absorption.

A sandwich ELISA was employed to quantify the amount of PSGL1/mIgG_{2b} in the supernatants of transfected COS cells. Typically, 5 culture flasks (260 ml flasks, Nunclon™) with COS cells at 70% confluence were transfected and incubated as described in materials and methods. Following an incubation period of 7 days in 10 ml of AIM V medium per flask, the medium was collected. The concentration of fusion protein in the supernatant from such a transfection, as well as in different volumes of supernatant following absorption on 100 µl gel slurry of anti-mouse IgG agarose beads (corresponding to 50 µl packed beads) was determined by an ELISA calibrated with purified mouse IgG Fc fragments (Fig. 2). The concentration of PSGL1/mIgG_{2b} in the supernatants ranged from 150 to 200 ng/µl, and in this particular experiment it was approximately 160 ng/µl (Fig. 2 A, the non-absorbed column). The concentration of PSGL1/mIgG_{2b} remaining in 2, 4 and 8 ml of supernatant following absorption on 50 µl packed anti-mouse IgG agarose beads was 32, 89 and 117 ng/µl, respectively. This corresponds to 260, 290 and 360 ng of PSGL1/mIgG_{2b} being absorbed onto 50 µl packed anti-mouse IgG agarose beads from 2, 4 and 8 ml of supernatant, respectively. Western blot analysis with the *B. simplicifolia* IB₄ lectin revealed that 50 µl of packed beads could absorb out the PSGL1/mIgG_{2b} fusion protein from 1 ml supernatant to below detectability and from 2 ml to barely detectable levels (not shown).

The absorption capacity of immobilized, Galα1,3Gal-substituted

PSGL1/mIgG_{2b}. Twenty ml of supernatant from COS cells transfected with the PSGL1/mIgG_{2b} plasmid alone or together with the porcine α1,3GT cDNA, were mixed with 500 µl gel slurry of anti-mouse IgG agarose beads each. Following extensive washing the beads were aliquoted such that 100 µl of gel slurry (50 µl packed beads) were mixed with 0.25, 0.5, 1.0, 2.0 and 4.0 ml of pooled, complement-depleted, human AB serum, and rolled head over tail at 4 °C for 4 hrs. Following absorption on PSGL1/mIgG_{2b} and Galα1,3Gal-substituted PSGL1/mIgG_{2b}, the serum was assayed for porcine endothelial cell cytotoxicity in the presence of rabbit complement using a 4 hr ⁵¹Cr release assay (Fig. 3). As shown in Fig. 3, 100 µl of beads carrying approximately 300 ng of PSGL1/mIgG_{2b} (see above) can reduce the cytotoxicity of 4 and 2 ml AB serum in each dilution step, and completely absorb the cytotoxicity present in 1 ml and less of human AB serum. Note that the same amount of non-Galα1,3Gal-substituted PSGL1/mIgG_{2b} reduces the cytotoxicity of 0.25 ml absorbed human AB serum only slightly (Fig. 3).

The effect of Gal α 1,3Gal-substituted PSGL1/mIgG_{2b} on complement-dependent porcine endothelial cell cytotoxicity and binding. To investigate the efficiency with which PSGL1/mIgG_{2b} could absorb individual human immunoglobulin classes, human IgG, IgM and IgA were purified from human AB serum by immuno-affinity chromatography on anti-human IgG, IgM and IgA agarose beads. Following its isolation IgA was passed through anti-IgG and IgM columns to remove traces of IgG and IgM. This procedure was performed for the other Ig classes as well. Ig fraction purity was checked by SDS-PAGE (Fig. 4). In concentrations found in normal serum, human IgG and IgM, but not IgA, were cytotoxic for PEC-A in the presence of rabbit complement (Fig. 5). The cytotoxicity residing in the IgG and IgM fractions was completely removed by absorption on Gal α 1,3Gal-substituted PSGL1/mIgG_{2b}. To investigate whether the lack of cytotoxicity exhibited by the IgA fraction was due to a lack of binding of human IgA antibodies to PEC-A, a cell ELISA was run with the same Ig fractions that was used in the cytotoxicity assay in order to detect bound IgG, IgM and IgA. Alkaline phosphatase-conjugated, class specific F(ab)₂ fragments were used as secondary antibodies. Eventhough the cytotoxicity of IgG and IgM was completely removed by absorption on Gal α 1,3Gal-substituted PSGL1/mIgG_{2b}, the binding was never reduced with more than 70 % (ranging from 30 to 70 %) for IgG, and never with more than 55 % (ranging from 10 to 55 %) for IgM (Fig. 5). Human IgA clearly bound to PEC-A, and the binding was only slightly reduced (not more than 29 %) following absorption on Gal α 1,3Gal-substituted PSGL1/mIgG_{2b}. The lack of cytotoxicity of the IgA fraction could therefore not be explained by an inability of the IgA fraction to bind PEC-A, but may be due to an inability to activate complement.

The effect of Gal α 1,3Gal-substituted PSGL1/mIgG_{2b} on ADCC of porcine endothelial cells. Several assays have been performed under serum-free conditions where PEC-A have had an intermediate sensitivity to direct killing by freshly isolated PBMC when compared to K562 and Raji cells; K562 being sensitive and Raji non-sensitive to killing by human NK cells (Fig. 6 A). In the presence of 10 % human, complement-inactivated AB serum, the killing rate was almost doubled supporting an ADCC effect *in vitro* (Fig. 6 B) in agreement with previously published data (30). However, if the serum is absorbed with the Gal α 1,3Gal-substituted PSGL1/mIgG_{2b} under conditions known to remove all PEC-A cytotoxic antibodies (see above), the killing rate decreases to levels slightly lower than those seen under serum-free conditions. On the other hand, the PSGL1/mIgG_{2b} fusion protein itself without Gal α 1,3Gal epitopes could not absorb out what caused the increased killing rate in the presence of human AB serum (Fig. 6 B). These data support the notion

that anti-pig antibodies with Gal α 1,3Gal specificity can support an antibody-dependent cell-mediated cytotoxicity *in vitro*, and that the Gal α 1,3Gal-substituted PSGL1/mIgG_{2b} fusion protein can effectively remove these antibodies just as it effectively removes the complement-fixing cytotoxic anti-pig antibodies.

Description of the drawings

Fig. 1. Six percent SDS-PAGE of proteins isolated from supernatants of COS cells transfected with vector alone (CDM8), PSGL1/mIgG_{2b}, or PSGL1/mIgG_{2b} and porcine α 1,3GT expression plasmids. Anti-mouse IgG agarose beads were used for immunoaffinity purification of fusion proteins. Following extensive washing, the beads were boiled in sample buffer under reducing and non-reducing conditions to release absorbed proteins. Gels run in parallel were either silver stained, or used for electrophoretic transfer of separated proteins onto nitrocellulose membranes. These were subsequently probed with peroxidase-conjugated *Bandeireia simplicifolia* isolectin B₄ lectin and visualized by chemiluminescence to detect Gal α 1,3Gal epitopes on immunopurified proteins. The gel migration length of molecular weight proteins of 220, 97 and 66 kDa is indicated on the left handside.

Fig. 2. Quantification by anti-mouse IgG Fc ELISA of the PSGL1/mIgG_{2b} fusion protein concentration in increasing volumes of transfected COS cell supernatants before and after absorption on 50 μ l of anti-mouse IgG agarose beads. Triplicate samples were analyzed.

Fig. 3. Antibody-dependent, complement-mediated PEC-A cell cytotoxicity by different volumes of human AB serum following absorption on 50 μ l of anti-mouse IgG agarose beads carrying approximately 300 ng of Gal α 1,3Gal- or non-substituted PSGL1/mIgG_{2b}, as estimated in a ⁵¹Cr-release assay.

Fig. 4. Ten percent SDS-PAGE of immunoaffinity purified human IgG, IgM, and IgA. Four micrograms of each sample were run under reducing and non-reducing conditions, and proteins were visualized by silver staining. The gel migration length of molecular weight proteins of 220, 97, 66, 46 and 30 kDa is indicated on the left handside.

Fig. 5. The antibody-dependent, complement-mediated PEC-A cell cytotoxicity of immunoaffinity purified human IgG, IgM and IgA before and after absorption on Gal α 1,3Gal-substituted PSGL1/mIgG_{2b} was investigated by ⁵¹Cr-release assays (right handside Y-axis; % killing). The PEC-A cell binding of immunoaffinity puri-

fied IgG, IgM and IgA before and after absorption on Gal α 1,3Gal-substituted PSGL1/mIgG_{2b} was estimated in a cell ELISA (left handside Y-axis; O.D. at 405 nm). The two assays were run in parallel on absorbed and non-absorbed Ig fractions. The concentration of the different Ig classes were chosen to be approximately half maximum of what is normally found in human serum.

Fig. 6. The direct cytotoxic effect (serum-free conditions) of human PBMC on K562, Raji and PEC-A cells, and the potentiating effect on killing by the addition of heat-inactivated, human AB serum, was investigated in a 4 hr ⁵¹Cr-release assay (graph A). The effect of heat-inactivated, human AB serum on antibody-dependent cellular cytotoxicity of PEC-A cells was studied in a 4 hr ⁵¹Cr-release assay before and after absorption on Gal α 1,3Gal- and non-substituted PSGL1/mIgG_{2b}, respectively (graph B).

DISCUSSION

Three major avenues of research have been followed in order to develop strategies to prevent HAR. Methods (i) to remove or neutralize anti-pig antibodies, (ii) to interfere with complement activation, and (iii) to remove or modify Gal α 1,3Gal determinants, have been tested in assays *in vitro*, in *ex vivo* organ perfusions and in pig to primate transplantations.

Pretransplant pig organ perfusion (46), plasmapheresis (13, 14, 20), immunoabsorption (13, 14, 17), and absorption on synthetically made Gal α 1,3Gal-containing oligosaccharides coupled to a solid phase (22) have been used to remove anti-pig antibodies. Recently, oligosaccharides derived from pig gastric mucin were shown to efficiently remove anti-Gal α 1,3Gal-specific antibodies and thereby anti-pig cytotoxicity (24). Injection of free saccharides to block anti-pig antibodies in the circulation is an alternative route used to prevent HAR. Intravenous carbohydrate therapy was used successfully in an ABO incompatible, heterotopic cardiac allograft model in the baboon (47) and in neonates with ABO hemolytic disease (48). No deleterious effects were noticed in recipient baboons upon injection of free blood group trisaccharides at rates corresponding to 500 mg/hr following a bolus injection of 4 g of the trisaccharide (47). Free Gal α 1,3Gal-containing saccharides have been used *in vitro* to neutralize anti-pig antibodies (23), but synthetic Gal α 1,3Gal-containing oligosaccharides in sufficient quantities for use in pig to primate xenotransplantation models are not yet available. However, high concentrations of melibiose (Gal α 1,6Glc) or arabinogalactan, a naturally occurring plant polysaccharide containing non-reducing α -D-galactose, have been used to prevent the toxic effect of ba-

boon serum on porcine PK15 cells *in vitro* and to prolong pig cardiac xenograft survival in transplanted baboons (49). Lethal toxic effects of the carbohydrate were observed in this study (49). Peptides mimicking the Gal α 1,3Gal structure (50, 51) and murine monoclonal anti-idiotypic antibodies directed against common idiotopes on naturally occurring human anti-pig antibodies (52), are new reagents that may be used to absorb or block anti-pig antibodies. In addition, anti- μ chain antibodies have been used in a guinea pig to rat xenotransplantation model to deplete xenoreactive IgM antibodies *in vivo* (15, 16) - a strategy that warrants further investigation in the pig to primate combination.

Cobra venom factor (20) and soluble complement receptor type 1 (21) have been used to perturb complement-activation and thereby to prolong xenograft survival in pig to non-human primate transplantations. Transgenic pigs have been made in which cDNAs encoding human sequences of the species-restricted complement-regulatory proteins, CD55 and CD59, are expressed in porcine endothelium (53-55). Organs from transgenic pigs have been shown to be less prone to HAR following organ perfusion with human blood or transplantation into non-human primates (56-59).

The most recent approach to prevent HAR involves modulating the expression of Gal α 1,3Gal epitopes on porcine endothelium. Sandrin and McKenzie suggested and proved that the expression of a glycosyltransferase, such as an α 1,2 FT, competing with the endogenous α 1,3 galactosyltransferase for the same precursor carbohydrate prevented expression of Gal α 1,3Gal epitopes upon transfection into porcine LLC-PK1 cells (26). As a result these cells became less sensitive to antibody-mediated, complement-dependent lysis (26). Recently, two independent groups have reported the making of transgenic pigs expressing this blood group H α 1,2 FT, but no data have been reported on the sensitivity of the endothelium derived from these pigs to antibody-mediated cytotoxicity (27, 28).

We describe the construction and production of a novel, efficient absorber that can be produced cheaply, in large quantities, and which can be easily coupled to a solid phase facilitating extracorporeal immunoabsorption of human blood to remove anti-pig antibodies before pig organ xenografting. To make possible the production of a highly glycosylated, recombinant protein carrying large amounts of Gal α 1,3Gal epitopes (Fig. 1), we made a chimeric protein by fusing the cDNA encoding the extracellular part of a membrane-anchored mucin, PSGL-1 (40), with the Fc part of mouse IgG, and coexpressed this with porcine α 1,3 GT. Mucins are the main

constituents of mucus, which is the gel covering all mucous membranes. The physical characteristics of mucins are due to their high content of O-linked carbohydrates (usually more than 60 % of the MW). Several cell membrane-anchored proteins containing mucin-type domains have been characterized: CD34, CD43, GlyCAM-1, PSGL-1, MAdCAM, CD96, CD45, and RBC glycoporphorins among others (43, 44). This group of proteins has received a lot of attention since many of them have been shown to carry the carbohydrate ligands of the selectin family of adhesion molecules; CD34, MAdCAM and GlyCAM-1 carry the carbohydrate epitope recognized by L-selectin and PSGL-1 carries epitopes recognized by both E- and P-selectin. Two reasons made us choose the PSGL-1 extracellular part as a fusion partner. First, it has one of the longest mucin-domains known in this family of proteins and may therefore be expected to carry most O-linked carbohydrates (44, 60). Second, PSGL-1 is not only a functional ligand for E-selectin, but is the hitherto only identified protein scaffold known to present sialyl-Le^x to P-selectin (61). So, if one were to make a fusion protein expressing sialyl-Le^x in order to inhibit P- and E-selectin-mediated adhesion, this should contain the PSGL-1 extracellular part. We are currently investigating the possibility of making a fusion protein expressing both Gal α 1,3Gal and sialyl-Le^x thereby exhibiting the characteristics of a dual inhibitor; one part neutralizing anti-pig antibodies and the other preventing E- and P-selectin-dependent rolling which may be a prerequisite for leukocyte extravasation in xenograft rejection.

In a xenotransplantation situation, where pig organs for transplantation would be readily available, the recipient could be prepared by extracorporeal immunoabsorption to remove anti-pig antibodies thereby preventing HAR. Alternative strategies could be chosen (see above), but a specific removal of anti-pig antibodies would leave the patient in a more favourable state with regard to the humoral immunity as compared to plasmapheresis and protein A absorption. Therefore, immunoabsorption media containing the epitopes recognized by the anti-pig antibodies themselves would be ideal. The production of Gal α 1,3Gal-containing di- and trisaccharides is laborious and costly, although a combination of organic and enzymatic synthesis have simplified the procedure and made it more cheap (34). Furthermore, it is not unlikely to expect the specificity of the human anti-pig antibody repertoire to be broader than just the Gal α 3Gal β 4GlcNAc epitope; *i.e.* recognition may be modulated by the core saccharide chain, neighbouring branching points and monosaccharides, as well as modifications such as sulfation. In this context it would be better to use a glycosylated, recombinant glycoprotein modified through the action of the porcine α 1,3 GT itself resulting in a more natural array of Gal α 1,3Gal-containing structures. Three

hundred ng of our fusion protein could completely remove the pig endothelial cell cytotoxic antibodies from 1 ml of human AB serum (Fig. 3), which should be compared to other studies where 1 g of oligosaccharide linked to a solid-phase was used to absorb 3 ml of human AB serum (23). However, direct comparisons are needed to state a difference in absorption capability. The fusion protein is most likely efficient due to a polyvalent expression of Gal α 1,3Gal-containing determinants and a possible expression of the epitope in a variety of structural contexts facilitating the absorption of a broader spectrum of the human anti-pig repertoire. Recent studies with pig gastric mucin-derived oligosaccharides indicate that polyvalency is important for absorption effectiveness (24).

Human IgG, IgM and IgA were purified from pooled human AB serum (Fig. 4) in order to investigate their binding to, and cytotoxicity for, porcine aortic endothelial cells, and to evaluate the Ig class-specific absorption efficacy of the fusion protein (Fig. 5). The PAEC cytotoxicity of human IgG at 8 mg/ml and IgM at 1 mg/ml was completely removed by Gal α 1,3Gal-substituted PSGL1/mIgG_{2b}. However, the binding was not completely abolished as estimated in a PAEC ELISA using aliquots of the same antibody preparations as in the cytotoxicity assay. Whether this represents residual IgG and IgM binding to epitopes other than Gal α 1,3Gal on PAEC - epitopes which are not able to induce an antibody-mediated cytotoxic effect, Fc receptor binding, or just a non-specific, non-functional binding, is not known at present. In our hands, purified IgA was not cytotoxic to PAEC although it clearly bound these cells (Fig. 5). This is in contrast with a previous study by Schaapherder et al, who demonstrated cytotoxicity by dimeric human IgA via the alternative pathway of complement activation (62). However, they used serum from human agammaglobulinemic donors as complement source, whereas we used rabbit serum (62). Furthermore, we saw no clear evidence of dimeric IgA in our IgA preparations (Fig. 4). In any case, IgA clearly binds PAEC and may be involved in pig xenograft rejection through other effector mechanisms mediated by, for instance, Fc receptor binding on macrophages and neutrophils (63, 64). This emphasizes the importance of using absorbers that remove all anti-pig Ig classes, something which protein A and anti- μ chain antibody absorptions do not.

Although anti-pig antibodies to a large extent rely on complement activation for full cytotoxic effect, there are additional effector mechanisms involving antibodies that may cause pig endothelial cell cytotoxicity. Antibody-dependent cellular cytotoxicity is such a mechanism where anti-pig antibodies with Gal α 1,3Gal specificity are important for cytotoxicity even in the absence of complement (30). Therefore, we

examined how absorbed and non-absorbed human AB serum contributed to PAEC cytotoxicity by human PBMC (Fig. 6). As shown in Fig. 6 human complement-depleted AB serum increased PEC-A cytotoxicity close to more than 10 % in 3 out of 4 effector to target ratios. This increased cytotoxicity contributed by human AB serum was completely removed by absorption on Gal α 1,3Gal-substituted PSGL1/mIgG_{2b} whereas absorption on PSGL1/mIgG_{2b} without Gal α 1,3Gal had no effect. This clearly demonstrates that Gal α 1,3Gal-specific antibodies not only contribute to ADCC against porcine endothelial cells, but are responsible for all of the enhanced cytotoxic effect seen upon addition of human AB serum to a human PBMC/PAEC mixed culture. If this ADCC effect is present also *in vivo*, strategies aiming at inhibiting complement activation may not be sufficient to prevent acute pig xenograft rejection.

We have presented the construction and production of a new and effective Gal α 1,3Gal-substituted, mucin domain-containing absorber that can be used in a pre-transplant extracorporeal immunoabsorption setting to remove anti-pig antibodies involved in antibody-dependent, complement- as well as cell-mediated cytotoxicity of pig endothelial cells. By stably transfecting cells to express the PSGL1/mIgG_{2b} and porcine α 1,3GT cDNAs, large amounts of fusion protein can be produced at low costs for testing in pig to primate xenotransplantation models.

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CLAIMS

1. An antigenic fusionprotein, which carries multiple Gal α 1,3Gal epitopes.
2. An antigenic fusionprotein according to claim 1, which is produced by a recombinant cell line.
3. An antigenic fusionprotein according to any one of claims 1 and 2, which is capable of binding preformed antibodies as well as antibodies produced as a response to a tissue or an organ originating from the species in which the Gal α 1,3Gal epitope is expressed, said species preferably being a species different from the antibody producing species.
4. An antigenic fusionprotein according to any one of the preceding claims, wherein the Gal α 1,3Gal epitopes have been made by an α 1,3 galactotransferase derived from a porcine species.
5. An antigenic fusionprotein according to any one of the preceding claims, which further comprises a part which mediates binding to selectins.
6. An antigenic fusionprotein according to claim 5, wherein the selectin is P-selectin.
7. An antigenic fusionprotein according to claim 5, wherein the part that mediates binding to selectin is a highly glycosylated protein, preferably a protein of mucin type.
8. An antigenic fusionprotein according to claim 7, wherein the part that mediates binding to selectin is the P-selectin glycoprotein ligand-1 (PSGL-1) or an essential part thereof.
9. An antigenic fusionprotein according to any one of the preceding claims, which further comprises a part which confers immunoglobulin properties.
10. An antigenic fusionprotein according to claim 9, wherein said part that confers immunoglobulin properties is an immunoglobulin or a part thereof, preferably IgG or a part thereof.

11. An antigenic fusionprotein according to claim 10, wherein said part that confers immunoglobulin properties is the Fc part of an immunoglobulin, preferably IgG, or an essential part thereof.
- 5 12. An antigenic fusionprotein according to claim 11, wherein said part that confers immunoglobulin properties is IgG_{2b}, preferably the Fc part thereof.
13. An antigenic fusionprotein according to any one of claims 9-12, wherein said part that confers immunoglobulin properties is of human origin.
- 10 14. An antigenic fusionprotein according to any one of the claims 9-12, wherein said part that confers immunoglobulin properties is of non-human origin and preferably is derived from mouse.
- 15 15. A cDNA molecule comprising a cDNA sequence coding for a fusionprotein according to any one of claims 1-14 or a derivative or variant thereof.
16. A vector which comprises a cDNA molecule according to claim 15 together with appropriate control and signal sequences.
- 20 17. A mammalian cell transfected with a vector according to claim 16.
18. A mammalian cell according to claim 17, which is a COS cell.
- 25 19. An absorber for use in the purification of blood plasma from antibodies raised against a foreign tissue or organ, which absorber comprises a fusion protein according to any one of claims 1-14.
- 30 20. A method for preventing a hyperacute rejection reaction in a patient who is to receive a xenotransplant, which method comprises withdrawal of plasma from the patient, bringing said plasma in contact with a fusionprotein according to any one of claims 1-14 to couple anti-pig antibodies thereto and thereafter reinfusing the plasma to the patient.

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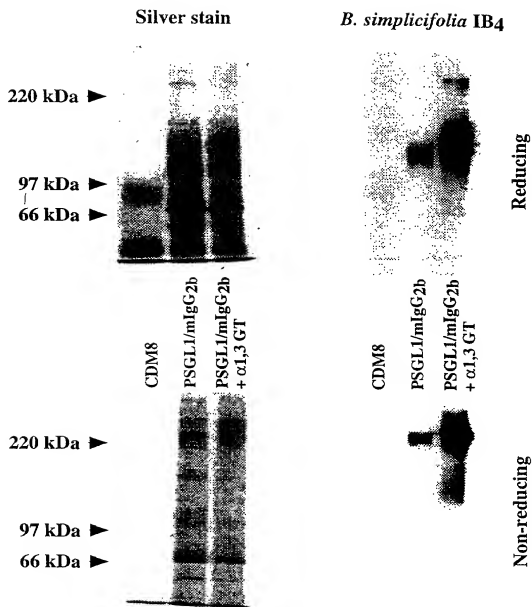


Fig. 1

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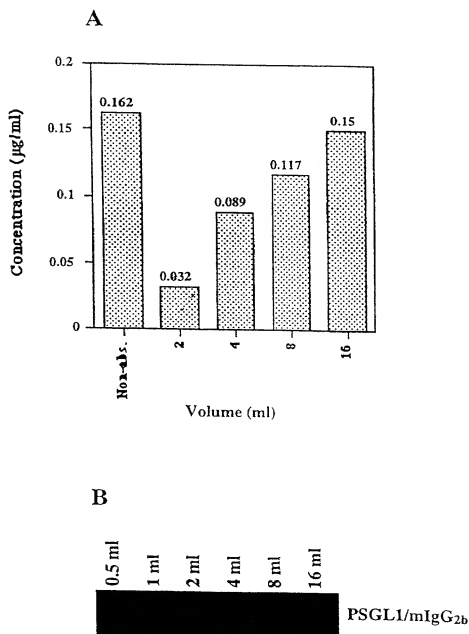


Fig. 2

Absorption capacity

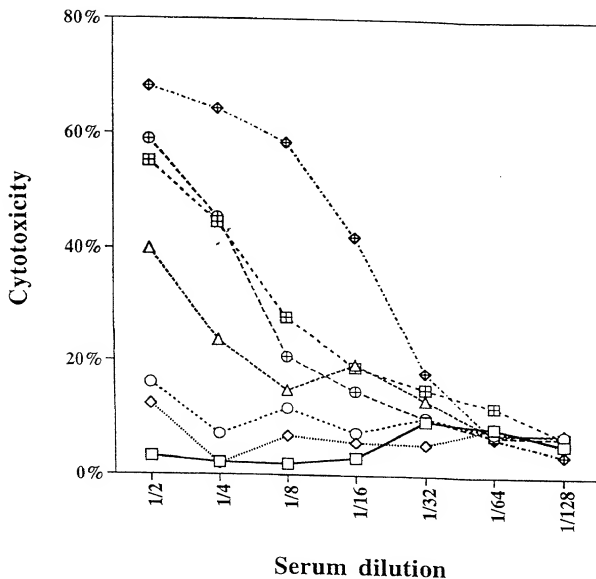


Fig. 3

- PSGL1/mIgG2b+GalT+0.25 ml AB serum
- ◇--- PSGL1/mIgG2b+GalT+0.5 ml AB serum
- PSGL1/mIgG2b+GalT+1 ml AB serum
- △--- PSGL1/mIgG2b+GalT+2 ml AB serum
- ⊠--- PSGL1/mIgG2b+GalT+4 ml AB serum
- ◆--- AB serum non-abs. 4 ml
- ⊕--- PSGL1/mIgG2b+0.25 ml AB serum

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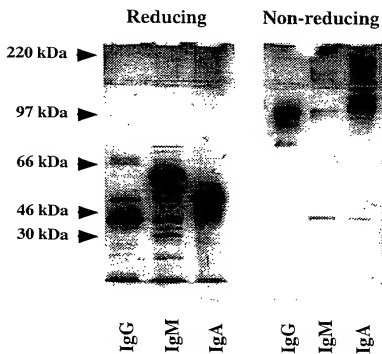


Fig. 4

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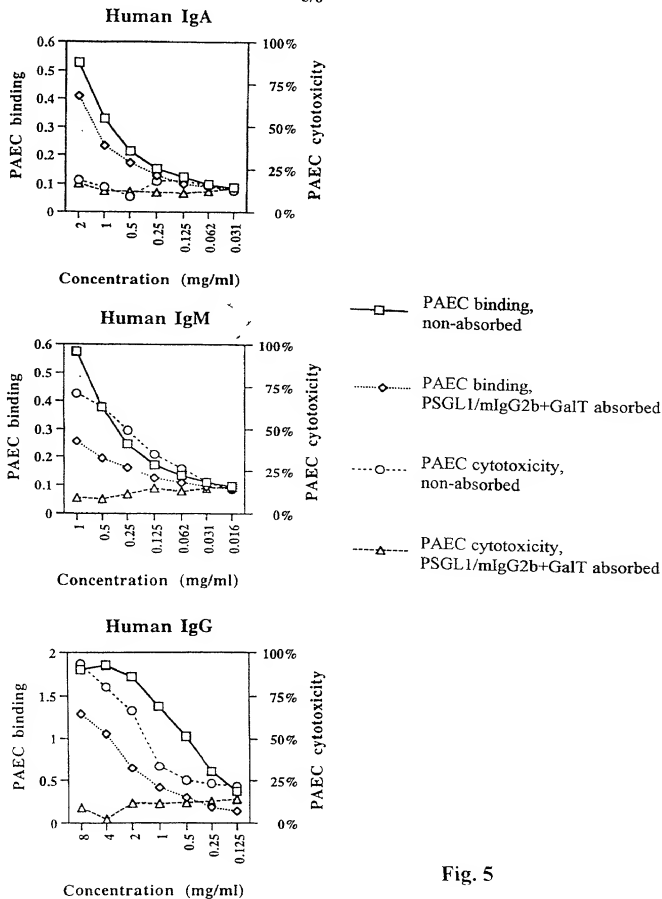
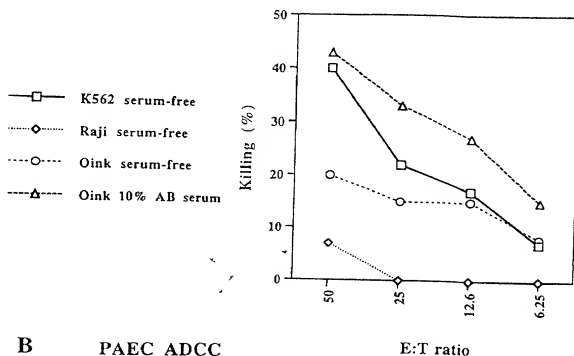


Fig. 5

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A ADCC



B PAEC ADCC

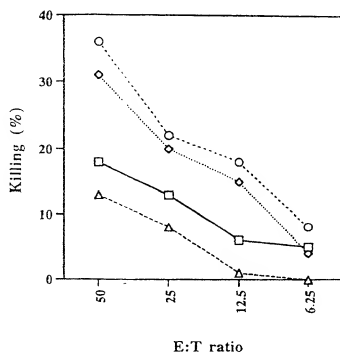
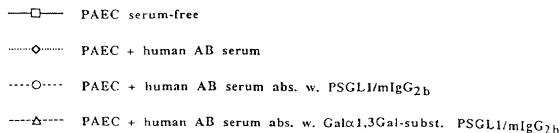


Fig. 6



COMBINED DECLARATION AND POWER OF ATTORNEY

As a below named inventor, I hereby declare that

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

ANTIGENIC FUSIONPROTEIN CARRYING GAL α 1,3GAL EPITOPES

the specification of which: *(check one)*

REGULAR OR DESIGN APPLICATION

☐ is attached hereto.

☐ was filed on _____ as application Serial No. _____
and was amended on _____ (if applicable).

PCT FILED APPLICATION ENTERING NATIONAL STAGE

☒ was described and claimed in International application No. PCT/SE98/00555 filed on 26 March 1998 and as amended on _____ (if any).

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, §1.56.

PRIORITY CLAIM

I hereby claim foreign priority benefits under 35 USC 119 of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed.

PRIOR FOREIGN APPLICATION(S)

Country	Application Number	Date of Filing (day, month, year)	Priority Claimed
Sweden	9701127-4	26 March 1997	yes

(Complete this part only if this is a continuing application.)

I hereby claim the benefit under 35 USC 120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of 35 USC 112, I acknowledge the duty to disclose information which is material to patentability as defined in Title 37 Code of Federal Regulations §1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application:

(Application Serial No.)

(Filing Date)

(Status—patented, pending, abandoned)

POWER OF ATTORNEY

- The undersigned hereby authorizes the U.S. attorney or agent named herein to accept and follow instructions from Albihs Patentbyra Stockholm AB as to any action to be taken in the Patent and Trademark Office regarding this application without direct communication between the U.S. attorney or agent and the undersigned. In the event of a change in the persons from whom instructions may be taken, the U.S. attorney or agent named herein will be so notified by the undersigned.

As a named inventor, I hereby appoint the following attorney(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith: **Robert J. PATCH, Reg. No. 17,355, Andrew J. PATCH, Reg. No. 32,925, Robert F. HARGEST, Reg. No. 25,590, Benoît CASTEL, Reg. No. 35,041, Eric JENSEN, Reg. No. 37,855, Thomas W. PERKINS, Reg. No. 33,027, and Roland E. LONG, Jr., Reg. No. 41,949, c/o YOUNG & THOMPSON, Second Floor, 745 South 23rd Street, Arlington, Virginia 22202.**

Address all telephone calls to Young & Thompson at 703/521-2297.

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

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